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WHAT IS DNA SEQUENCING



The term DNA Sequencing refers to is the process of determining the precise order of nucleotides (A/T/C/G) in a DNA molecule.



## WHAT IS DNA SEQUENCING ?

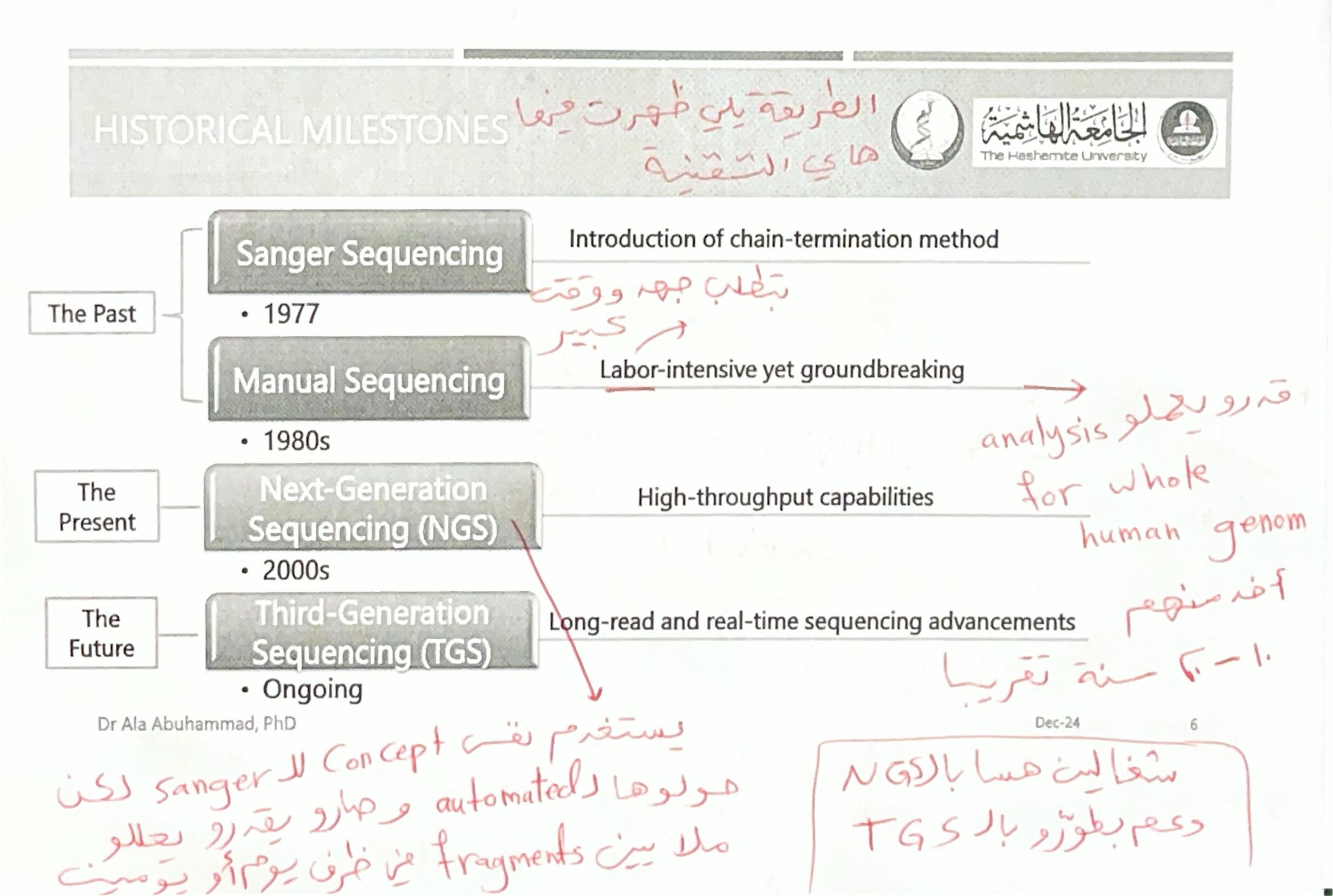


- This method reveals the sequence of the genetic code which provides the instructions for building and maintaining an organism.
- DNA Sequencing is fundamental in fields like genetic biology, biotechnology, medical research
- Also, DNA Sequencing allows scientists to study genetic variation, mutations, and evolutionary relationship.

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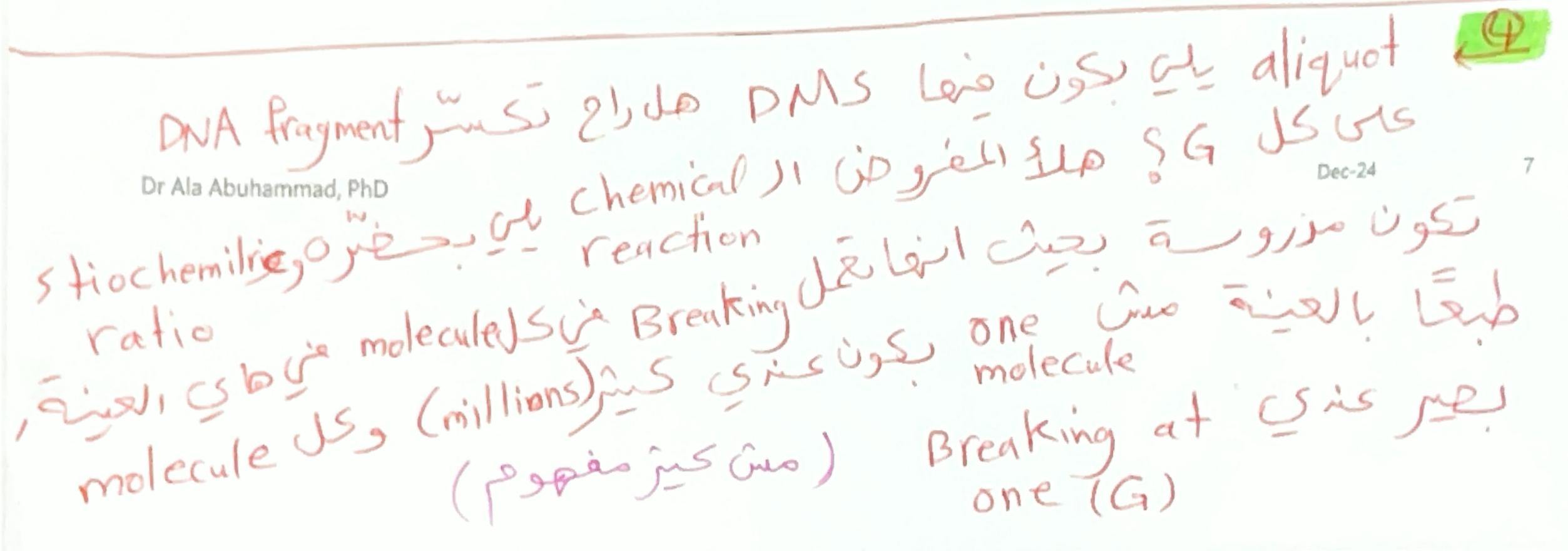
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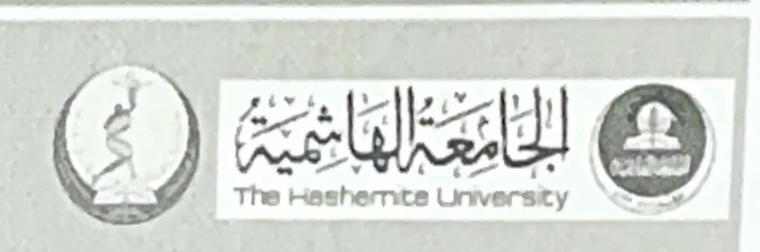
## HISTORICAL MILESTONES ...



- At the end of the 70s the first two rapid nucleic acid sequencing techniques were published almost simultaneously:
- 1. The Chemical Method of Allan Maxam and Walter Gilbert, and
- 2. The Enzymatic Chain Termination Method of Frederick Sanger, S. Nicklen, and A. R. Coulson.
  - In 1980, Sanger and Gilbert received the Nobel Prize in Chemistry for the development of DNA sequencing technology.
  - more popular
- Without changing the underlying concept of both methods, some improvements have been done over the years by applying different strategies, by developing various modifications and by automation.



## MAXAM-GILBERT CHEMICAL METHOD



- The Maxam-Gilbert chemical method starts by obtaining a set of double-stranded DNA molecules labelled at the 5' end with dNTPs having a radioactive isotope (usually 32P), by the action of polynucleotide kinase.
- Four aliquots of the same sample are then treated under different conditions. The chemical treatment generates breaks in a small proportion of one or two of the four nucleotides in each of the four reactions (G, A+G, C, C+T). The chemical agents used in each case are: Brea

5 end and i is labelling last selecte Jerial const fragment

) \* Formic acid (A+G) 4 aliquots legistics Sequencing by lost of Sample Hydrazine plus salts (C) (identical in the DNA sequence)

chemical reagent 11 pouls cience

- This generates a series of fragments, ranging in size from the radiolabelled end to the first "cut" site on each molecule.
- As the treatment is gentle, only one nucleotide per DNA molecule is cleaved, resulting in molecules of different sizes. The products of the four reactions are separated on a polyacrylamide gel. A radiographic plate is attached to observe the radioactive banding pattern formed.

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they will result: Chemical - 1 1000-24 in Breaking of the agents response tragment at a selective point

It can read & software view automatic ilsd & these bands, so they can gusse the DNA sequicing under each column MAXAM-GILBERT CHEMICAL METHOD An example of Maxam-Gilbert Sequencing Technique, showing specific cleavage of DNA backbone yielding different sized labelled DNA fragments. 1) Cittain single stranded DNA. denaturation ACTGACTGAA "ACTGACTGAA 2) Add a "P to 5" and 3) Cleave at specific nucleotides 4) Offerently sized DNA strands Electrophoresis through https://www.youtube.com/watch?v= high resolution acrylamide gate B5Di8PL4E0 Dr Ala Abuhammad, PhD E) Deduce DNA sequence (broken at certain size tube 15 is tragments Separationis re 219 get electrophin ve running replace MAXAM-GILBERT CHEMICAL METHOD Maxime Gilbert was a significant breakthrough The 1st widely adopted method for DNA sequencing.

It represents the 1st generation of DNA sequencing methods. This method allows the reading of about 100 sequence bases. however, it is less commonly used today, due to its complexity and the availability of more advanced methods. It has been largely replaced by faster and easier methods, like single sequencing or modern next generation sequencing (NGS) techniques. , they correspond to what fragment Julyan job lisa a pain color de comes pond to what fragment Julyan job lisa a pain color de coesa

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6 to build a complementary DNA strand using the 4 nucleotides + modefied mixture) we can be a modefied mixture) we can be a modefied mixture) deoxynecleotide Sanger Enzymatic Method The enzymatic Sanger chain termination method is based on the use of ddNTPs. As with the chemical method, four different reaction (P)mixtures are made. Each reaction mixture contains: DNA to be sequenced: plasmid with the "problem" DNA fragment or PCR product.

\* The four dNTPs: dATP, dCTP, dTTP and dGTP

\* DNA polymerase I

A radiotabelled or fluorescently labelled primer: a 17-20 base oligonucleotide complementary to the DNA fragment to be sequenced.

A ddNTP (at a low concentration): a dideoxynucleotide, which lacks a free 3' OH end, so that when incorporated into a DNA strand stops DNA elongation.

For Sanger sequencing ratio of dNTP: ddNTP ≈ 100:1

Fluotopho ddATP ddTTP ddGTP ddCTP Dideoxynucleotide (ddNTP) Deoxynucleotide (dNTP)

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Sanger Enzymatic Method

Cytosin

Sanger sequencing, also known as dideoxy sequencing or chain termination method, identifies the order of nucleotide bases in DNA. This method relies on chain termination by modified nucleotides called dideoxynucleotide triphosphates (ddNTPs).

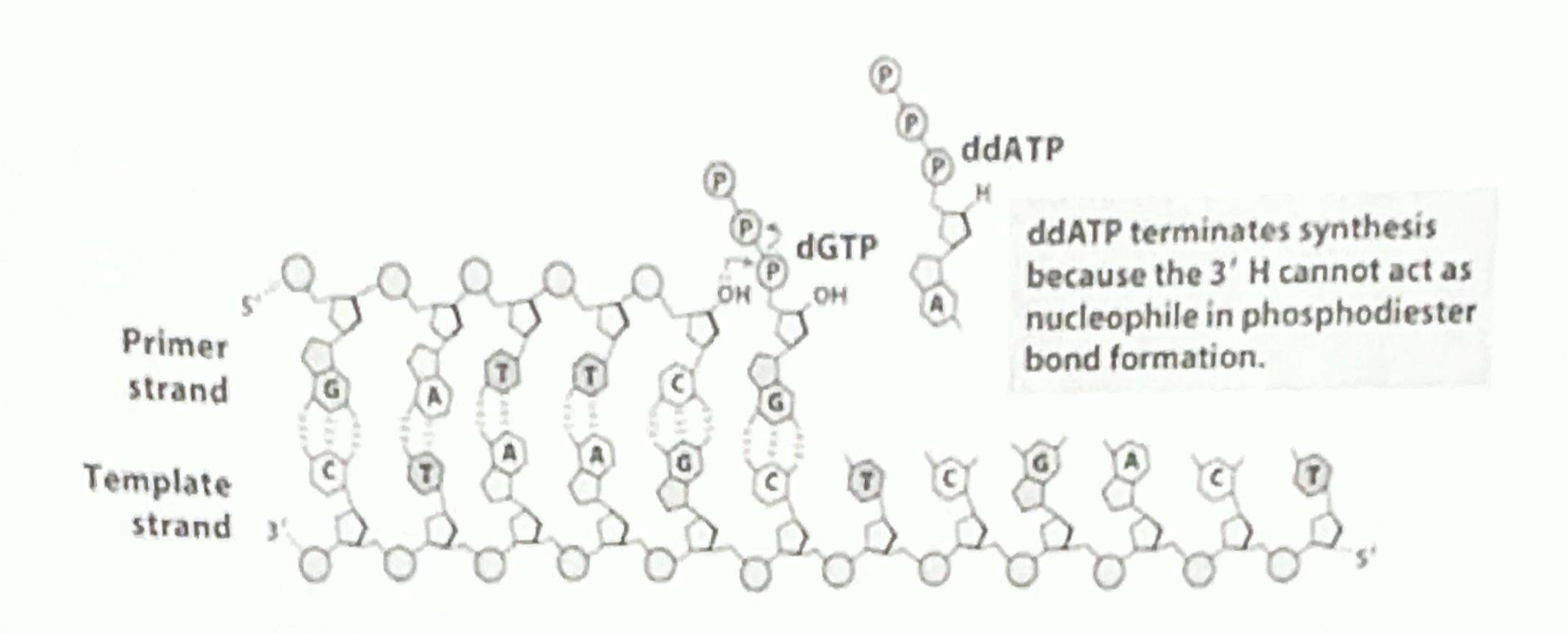
it was the first DNA sequencing method and remains highly regarded for its accuracy and the length of the reads it produces (~500-1000 bp).

The technique involves the incorporation of ddNTPs into a growing DNA chain during replication. These ddNTPs lack a 3' hydroxyl group, which prevents the addition of further nucleotides, thus terminating the chain. By incorporating fluorescent or radioactive labels, the terminated fragments can be detected and analyzed to determine the DNA sequence.

Sanger sequencing was crucial for the Human Genome Project, completed in 2003, marking a significant milestone in genomics. Today, it remains a fast and cost-effective method for reading small, targeted regions of the genome. Its applications include testing for known familial variants, validating NGS results, and single gene sequencing.

Dr Ala Abuhammad, PhD & 6H dr. A) leie ddNTP grlp golie
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another neoclotideine new den, PNA polymerase II de lie enzymatic chain termination





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#### Sanger Enzymatic Method 4 × PCR (+ one dideoxynucieotide) make multiple copies add to four attach a of a segment polymerase solutions primer denature dsDNA Use a using heat sequencing GAGTGAAGGT machine DNA sequence CTGACTTCGA electrophorese the grow complementary denaturate the **MINISTER** chains until termination dye four solutions grown chains TCG Separate with a gel 2000 NOOD Dr Ala Abuhammad, PhD - Automated part with Sanger's

## Sanger Enzymatic Method





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https://youtu.be/dVRB4CaLizc?feature=shared

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### Sanger Sequencing Enabled Genome Sequencin







1995

First bacterial genome (Haemophilus influenzae) sequenced

First draft of the human genome published

1995 First

2001 First

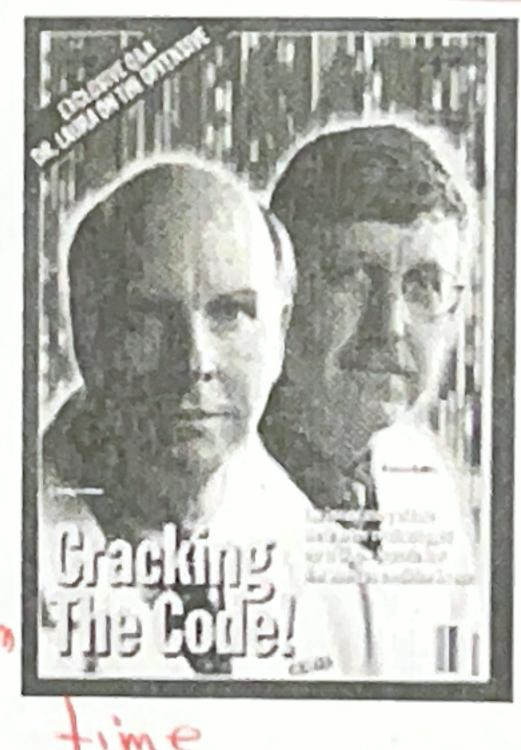
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## AUTOMATIC SEQUENCING

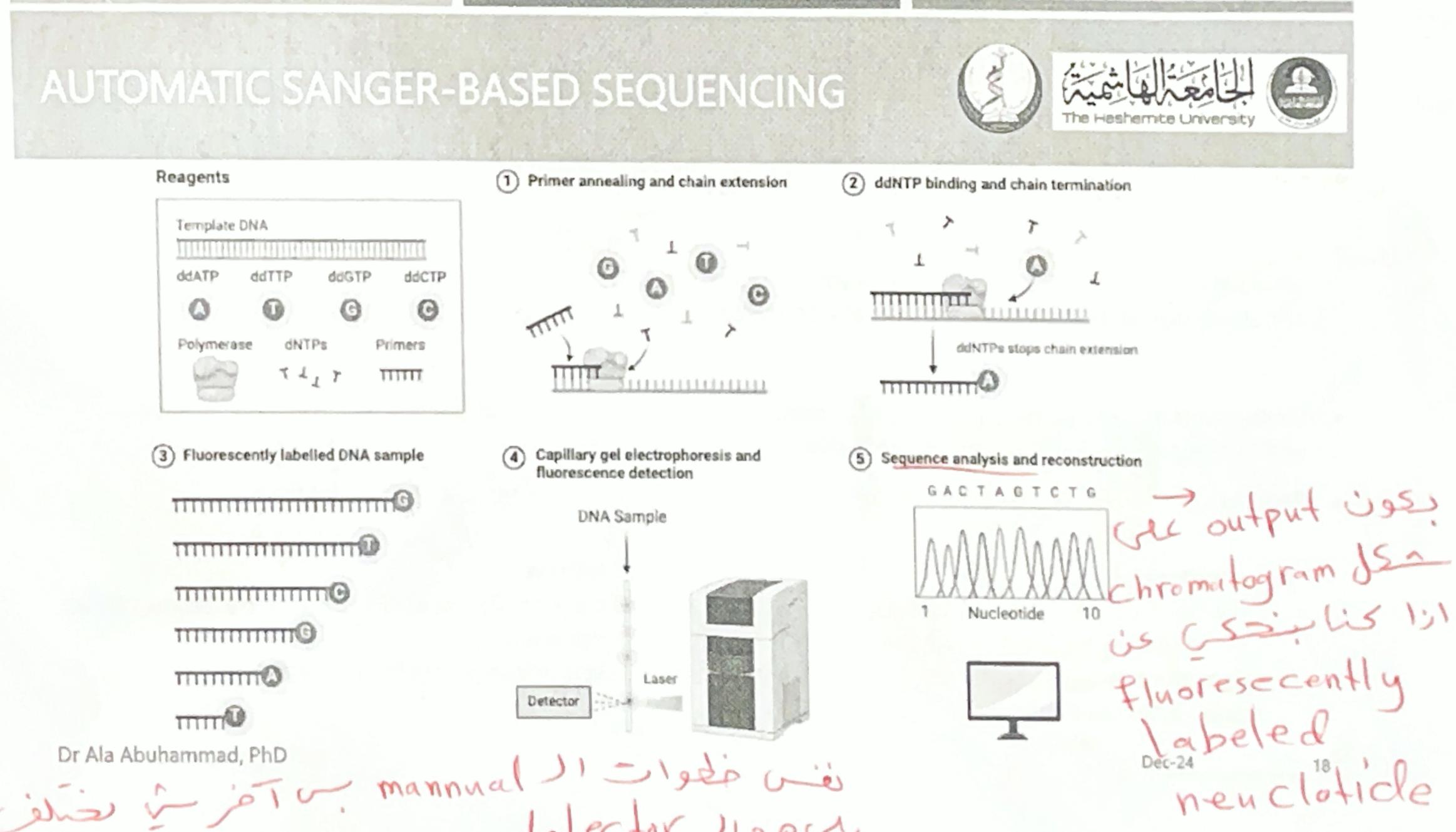


## 1. Automatic sanger-based sequencing

Aspect	Manual Sanger Sequencing	Automated Sanger Sequencing
Reading Method	User reads all four gel lanes from bottom to top.	Computer reads the bands in capillary gel.
Detection of ddNTP	Identified by the band's position in a specific lane (corresponding to ddNTP).	Laser excites fluorescent tags, and a computer detects the emitted light.
Fluorescence	No fluorescence; bands are detected using radioactive or UV light.	Fluorescent tags attached to ddNTPs emit distinct light signals.
Data Output	Manual interpretation of gel bands	Automated chromatogram with fluorescent peaks for each nucleotide.
Efficiency	Time-consuming and requires manual effort.	Faster, with less manual input and higher throughput.

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### AUTOMATIC SANGER-BASED SEQUENCING

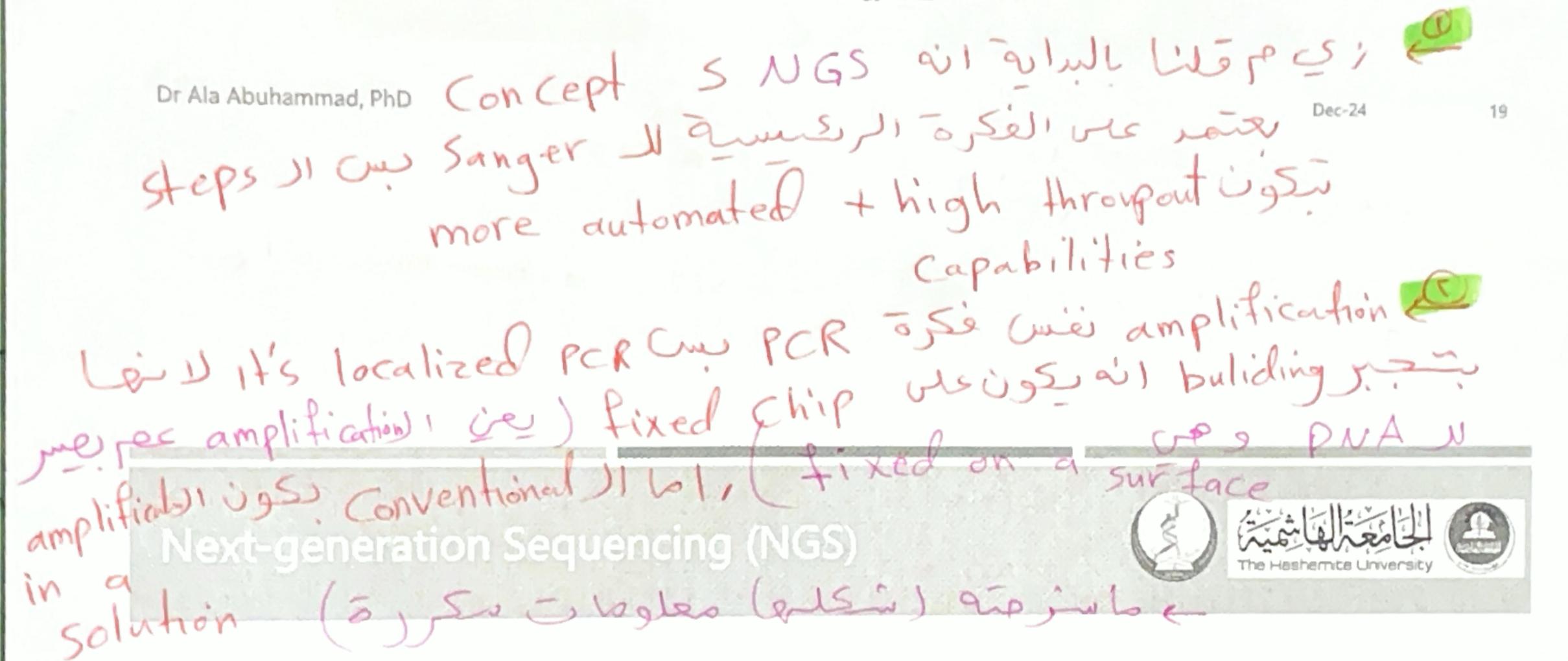


## AUTOMATIC SEQUENCING



### 2. Next-Generation Sequencing (NGS)

- Techniques based on the Sanger method are suitable for sequencing relatively small DNA fragments (up to 1000 bp), using cloning vectors such as plasmids or directly PCR products. However, for mass sequencing of larger fragments, such as the entire genome of an organism, Next generation sequencing (NGS) is used. These are a group of mass sequencing techniques.
  - Library Preparation: DNA is fragmented, and adaptors are attached to the fragments.
  - Amplification: The fragments are amplified using PCR.
  - Sequencing: The amplified fragments are read in parallel using platforms like Illumina or Ion Torrent.
  - Data Analysis: Bioinformatics tools interpret the sequences.
- Watch: <a href="https://www.youtube.com/watch?v=CZeN-lgjYCo">https://www.youtube.com/watch?v=CZeN-lgjYCo</a>



- NGS refers to advanced DNA sequencing technologies that allow for the rapid sequencing of large amounts of DNA. Unlike traditional Sanger sequencing, NGS is high-throughput, enabling the simultaneous sequencing of millions of fragments.
- Next-generation sequencing (NGS), also known as high-throughput sequencing, is the catch-all term used to describe a number of different modern sequencing technologies.
- Read lengths typically range, depending on the technology used, between 100 600 bp.

### Advantages

- High throughput and scalability
- Cost-effective for large-scale projects
- Capable of sequencing entire genomes, exomes, or transcriptomes

### Limitations

- Requires advanced computational resources for data analysis
- High (initial) costs for instruments

## Next-generation Sequencing (NGS)



- Key Steps in NGS
- 1. Sample Preparation
  - DNA is fragmented, and adapters are added to the ends for amplification and sequencing.
- 2. Library Preparation
  - Fragments are amplified on a solid surface (flow cell) using techniques like bridge amplification.
- 3. Sequencing by Synthesis (SBS)
  - DNA is synthesized base-by-base with fluorescently labeled nucleotides, and the sequence is recorded.
- 4. Data Analysis
  - The resulting short reads are aligned to a reference genome or assembled de novo.

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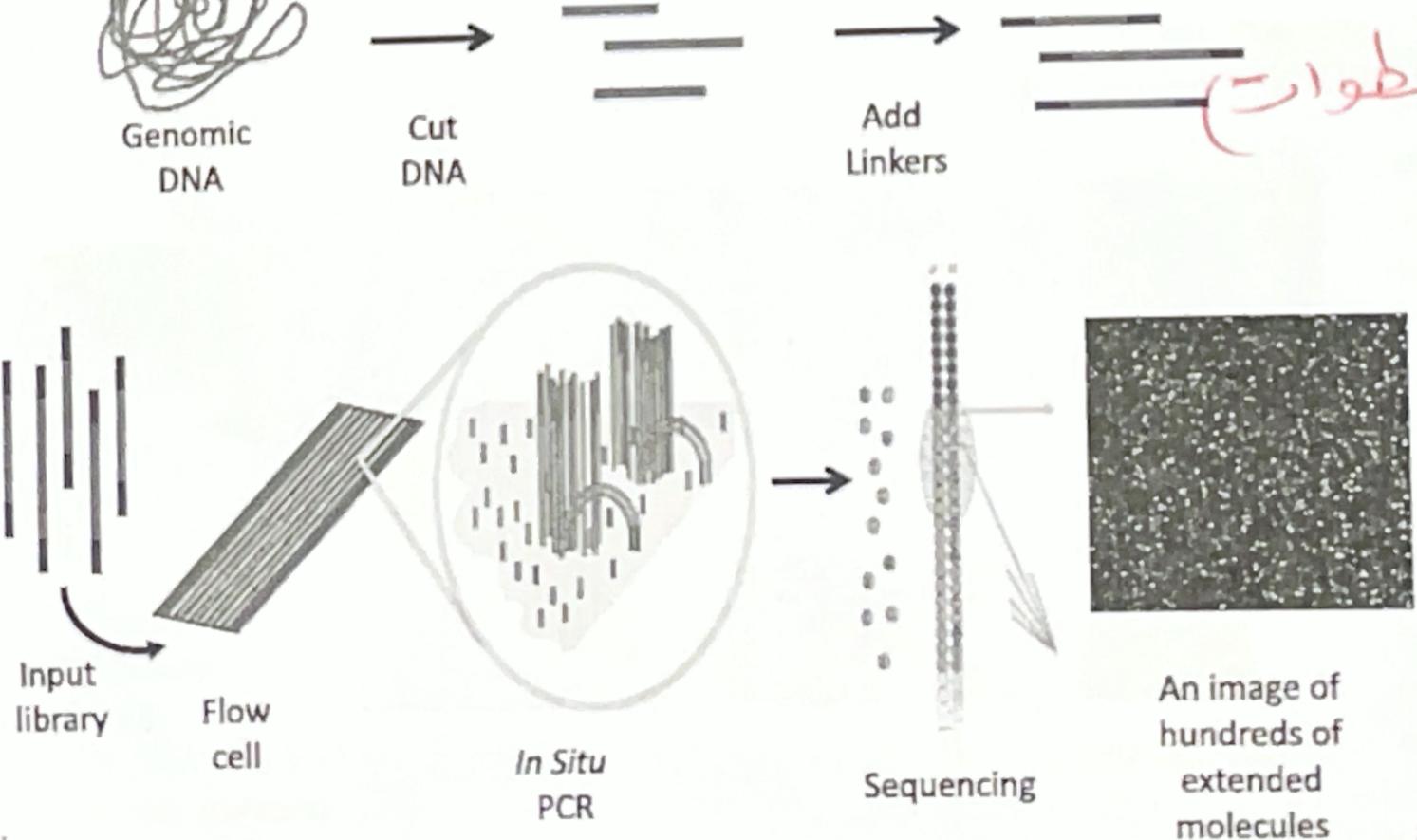
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Next-generation Sequencing (NGS)







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## Next-generation Sequencing (NGS)



- In Next-Generation Sequencing (NGS), DNA adaptors are short, synthetic double-stranded DNA sequences that are attached to the ends of DNA fragments during library preparation. These adaptors serve multiple essential roles in the sequencing process:
- Key Functions of DNA Adaptors in NGS:
- Facilitating Attachment to Sequencing Platforms:
  - Adaptors contain sequences that enable the DNA fragments to bind to the surface of a sequencing flow cell (e.g., in Illumina sequencing) or to beads (e.g., in Ion Torrent sequencing).
- Priming for Amplification and Sequencing:
  - They include sequences recognized by primers for PCR amplification, which generates sufficient copies of each fragment for sequencing.
  - During sequencing, the adaptors provide binding sites for sequencing primers.
- Indexing/Barcoding:
  - Adaptors often include index sequences (barcodes) that allow the multiplexing of multiple samples in a single sequencing run.
    These indices enable bioinformatics tools to differentiate reads originating from different samples.
- Compatibility with Sequencing Chemistry:
  - Adaptors are designed to be compatible with the sequencing platform's specific chemistry and workflow. For example, Illumina adaptors have regions that enable cluster formation on the flow cell.

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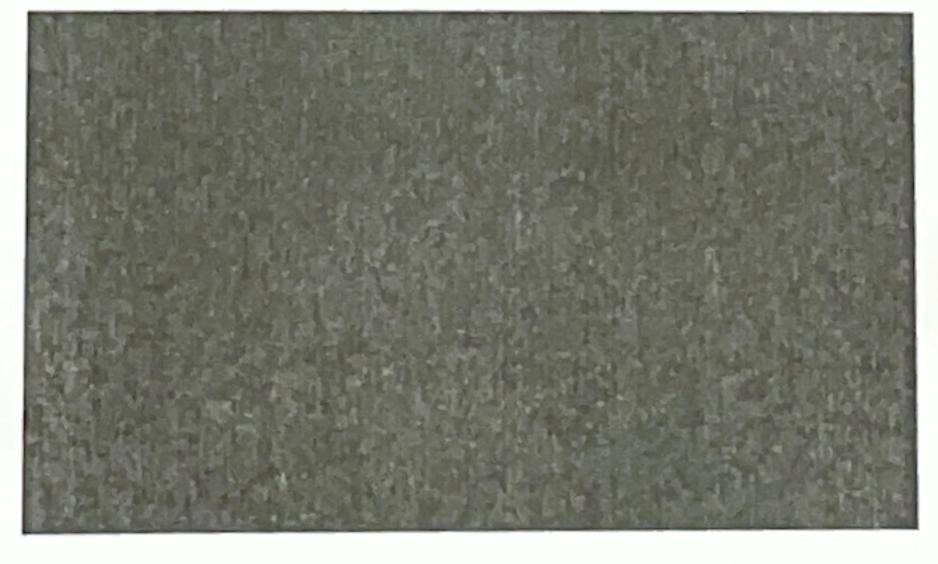
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### Next-generation Sequencing (NGS)



Illumina Sequencing is a widely-used next-generation sequencing (NGS) technology that allows for the high-throughput sequencing of DNA. It uses a method called sequencing by synthesis (SBS), which involves capturing DNA fragments on a solid surface and incorporating labeled nucleotides one base at a time.



https://www.youtube.com/watch?v=fCd6B5HRaZ8

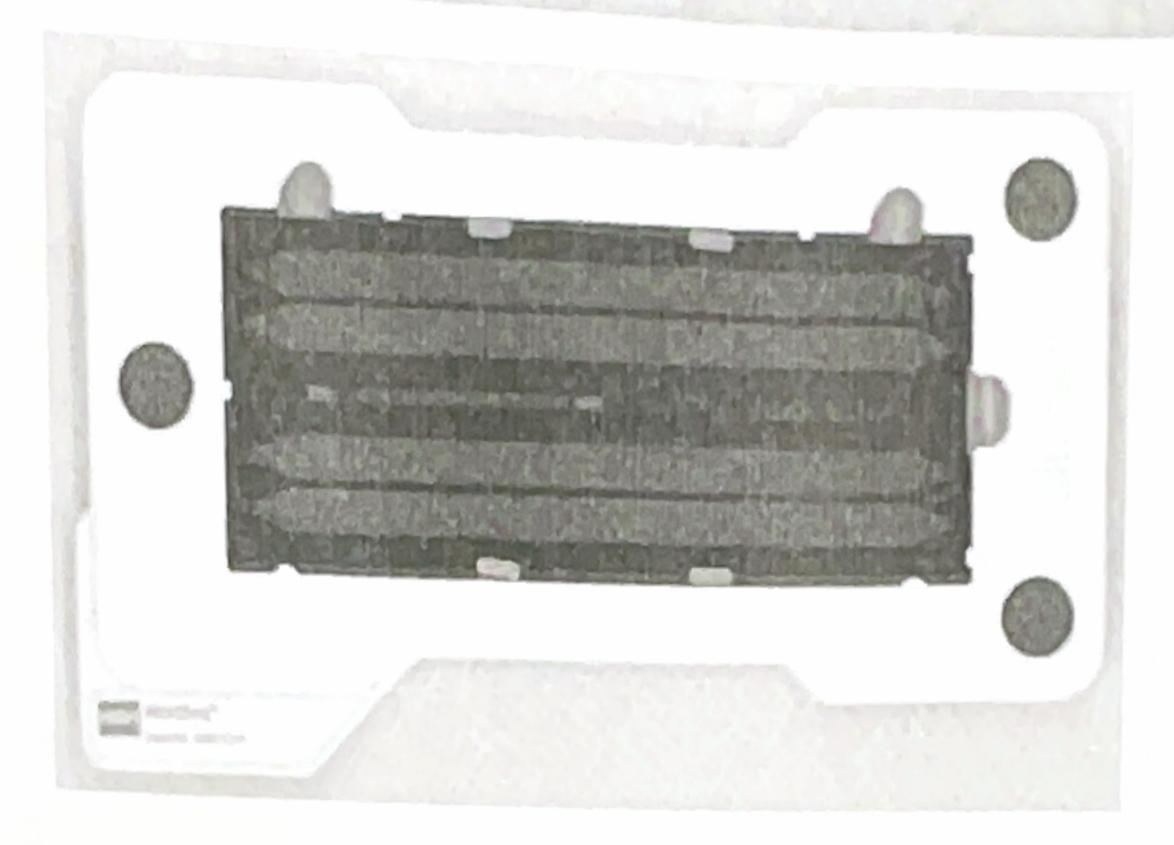
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## Illumina NextSeg Flow Cell





400,000,000 DNA fragments X 300 nucleotides

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120,000,000,000 nucleotides in 29 hours

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400 billion live large or end chip cla chip DNA fragment nucleotids vy length 11 fragments

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Illumina Sequencing Benchmarks



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NextSeq 500



VS.

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The Human Genome Project

1 Technician 29 hours \$4000 120,000,000,000 nucleotides Human genome X 40

>1000 Scientists 13 years (1990-2003) \$3,000,000,000 24,000,000,000 nucleotides = (1) Human genome X 8 The Sanger method remained the primary technique for the majority of sequencing, while newer techniques, such as

pyrosequencing and sequencing by synthesis (as seen in Illumina technology), started to emerge later and contributed to genome sequencing efforts in subsequent projects.

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### The Human Genome Project (HGP)







## (55/5/5/5)

- The Human Genome Project (1990–2003) was a landmark international research initiative aimed at mapping and sequencing the entire human genome. The goal was to identify all the genes in human DNA and understand their function, providing a foundation for advances in medicine and biology.
- Scope: Sequenced 3 billion DNA base pairs of the human genome.
- Output: Identified approximately 20,000–25,000 genes.
- Collaboration: Involved scientists from multiple countries and institutions.
- Impact: Revolutionized understanding of genetic diseases, drug discovery, and personalized medicine.

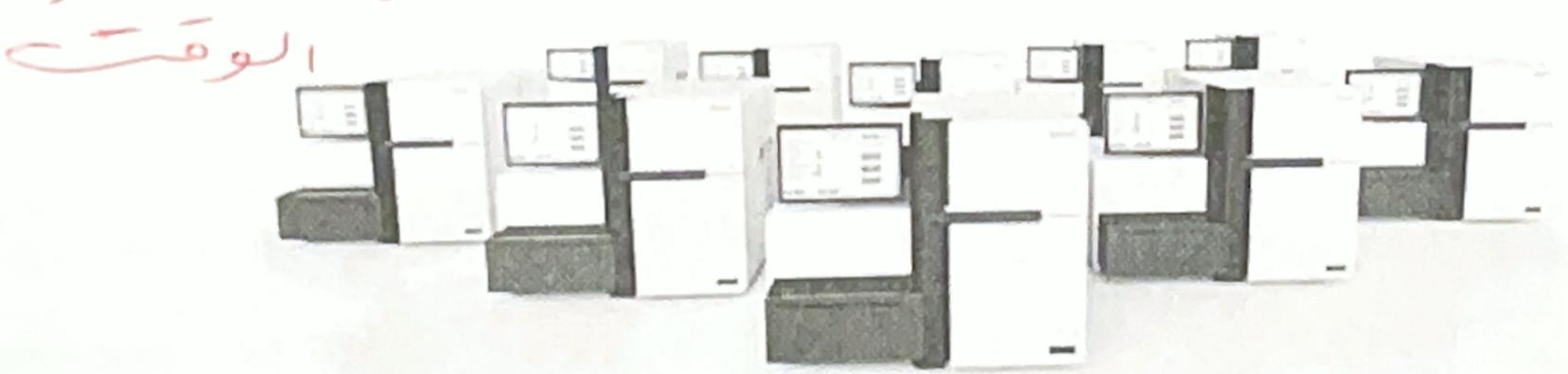
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# Illumina Sequencing Is Scalable نقه را شتفل علی کار دو من محار سفا





HiSeq X Ten: Single device has 15X capacity of the NextSeq 500

Cost for sequencing of a human genome (at 40X): ~\$1000

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## AUTOMATIC SEQUENCING



### 3. Pyrosequencing

- automated sequencing technique is pyrosequencing. It allows the determination of DNA sequences on a large scale, using luminescence. This technique requires:
- eDNA to be sequenced: single-stranded
- m Primer
- a dNTPs
- DNA polymerase
- 3 enzymes: sulphurylase (and the substrate adenosine 5'-phosphosulphate or APS), luciferase (and the substrate luciferin) and apyrase.
- The process occurs in successive three-step cycles. In the first step, one of the 4 dNTPs is added. If it is complementary to the base of the template strand to be copied, it is incorporated into the strand being extended by DNA polymerase, releasing a PPi. This reacts with APS to give ATP, which is used to oxidise luciferase, a process in which it emits light. If the dNTP that has been added to the reaction medium is not complementary, it is degraded by apyrase before the next dNTP is added. The emitted light is captured by a camera. The results are plotted as a pyrogram.
- This technique allows the sequencing of small and medium-sized DNA fragments (100 500 BP).

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https://www.youtube.com/watch?v=pollGc5uj7A

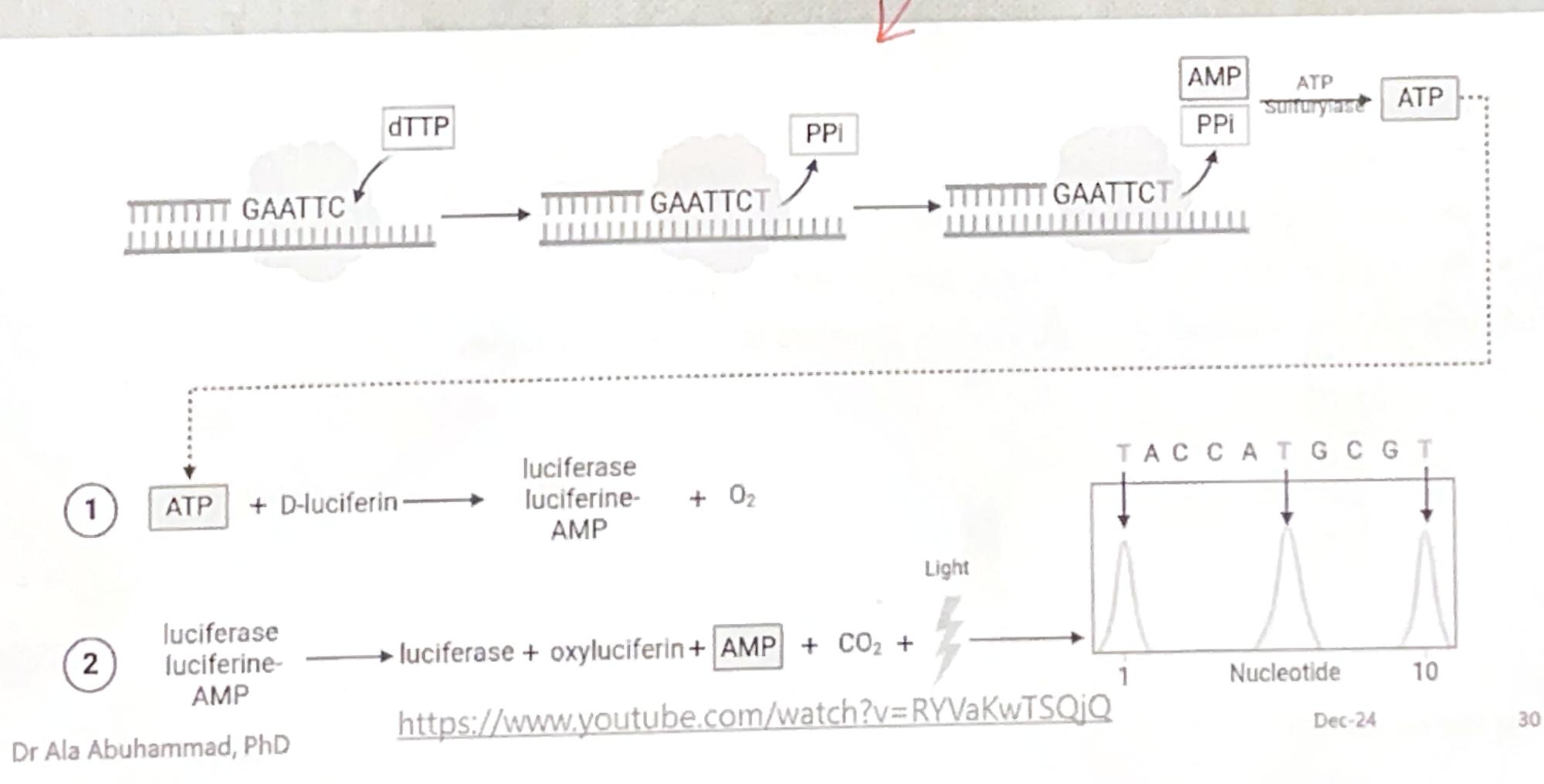
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### PYROSEQUENCING





## THIRD-GENERATION SEQUENCING (TGS)







Wins fragmentation Los 1 2 to 26

- Key Features:
  - Real-time sequencing of single molecules.
  - Long-read capabilities.
- Technologies:
  - PacBio (SMRT): Long reads, high accuracy.
  - Oxford Nanopore: Portable and versatile.

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## APPLICATIONS IN DRUG DISCOVERY



- Key Areas:
  - Target Identification: Detect mutations or gene variations linked to disease.
  - Biomarker Discovery: Identify genetic signatures for drug response.
  - Pharmacogenomics: Tailoring treatments based on genetic profiles.
- Example: Using sequencing to study tumor genomes for targeted therapies.

## CHALLENGES AND FUTURE DIRECTIONS



- Current Challenges:
  - Cost and data management for large-scale projects.
  - Ethical concerns in genomics.
- Future Innovations:
  - Integrating Al for data analysis.
  - Expanding accessibility to low-resource settings.
  - Improving accuracy for clinical applications.

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study hard, you are almost done





THE END





Any questions? ©

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